Product Datasheet

Biological Description

Specificity	Phospho-CREB (Ser133) Antibody [K11M21] detects endogenous levels of CREB only when phosphorylated at serine 133. The antibody also detects the phosphorylated form of the CREB-related protein, ATF-1.
Background	Phospho-CREB (Ser133) is the phosphorylated form of the cAMP response element-binding protein (CREB), a bZIP transcription factor essential for gene regulation in response to extracellular signals. Structurally, CREB contains a basic region-leucine zipper (bZIP) domain for DNA binding and dimerization, a kinase-inducible domain (KID) where Ser133 resides, and Q1/Q2 domains for transcriptional activation. Phosphorylation at Ser133, primarily by PKA, MSK1/2, and CaMK, enhances CREB's transcriptional activity by promoting interactions with co-activators CBP and p300. This modification integrates multiple signaling pathways, regulating genes involved in neuronal plasticity, metabolism, survival, and immune responses. Widely expressed in various tissues, especially the brain, phospho-CREB (Ser133) plays a pivotal role in learning, memory, and adaptive cellular responses, making it a key target in neurological and metabolic research.

Usage Information

Application	WB, IHC, IF, FCM, ChIP	Dilution	
Reactivity	Human, Mouse, Rat		
Source	Rabbit Monoclonal Antibody	MW	36 kDa
Storage Buffer	PBS, pH 7.2+50% Glycerol+0.05% BSA+0.01% NaN3		
Storage (from the date of receipt)	-20°C (avoid freeze-thaw cycles), 2 years		

WB	IHC	IF	FCM	CHIP
1:1000	1:400 - 1:1600	1:400 - 1:1600	1:400 - 1:1600	1:50

Experimental Methods

WB

Experimental Protocol:

Sample preparation
1. Tissue: Lyse the tissue sample by adding an appropriate volume of ice-cold **RIPA/Nuclear Lysis Buffer (containing Protease Inhibitor Cocktail)**, and homogenize the tissue at a low temperature or lyse it by sonication on ice, then incubate on ice for 30 minutes.
2. Adherent cell: Aspirate the culture medium and transfer the cells into an EP tube. Wash the cells with ice-cold PBS twice. Add an appropriate volume of **RIPA/Nuclear Lysis Buffer (containing Protease Inhibitor Cocktail)**, sonicate to lyse the cells, and incubate on ice for 30 minutes.
3. Suspension cell: Transfer the culture medium to a pre-cooled centrifuge tube. Centrifuge and aspirate the supernatant. Wash the cells with ice-cold PBS twice.Add an appropriate volume of **RIPA/Nuclear Lysis Buffer (containing Protease Inhibitor Cocktail)**, sonicate to lyse the cells, and incubate on ice for 30 minutes.
4. Place the lysate into a pre-cooled microcentrifuge tube. Centrifuge at 4°C for 15 min. Collect the supernatant;
5. Remove a small volume of lysate to determine the protein concentration;
6. Combine the lysate with protein loading buffer. Boil 20 µL sample under 95-100°C for 5 min. Centrifuge for 5 min after cool down on ice.

Electrophoretic separation
1. According to the concentration of extracted protein, load appropriate amount of protein sample and marker onto SDS-PAGE gels for electrophoresis. Recommended separating gel (lower gel) concentration: 10%. Reference Table for Selecting SDS-PAGE Separation Gel Concentrations
2. Power up 80V for 30 minutes. Then the power supply is adjusted (110 V~150 V), the Marker is observed, and the electrophoresis can be stopped when the indicator band of the predyed protein Marker where the protein is located is properly separated. (Note that the current should not be too large when electrophoresis, too large current (more than 150 mA) will cause the temperature to rise, affecting the result of running glue. If high currents cannot be avoided, an ice bath can be used to cool the bath.)

Transfer membrane
1. Take out the converter, soak the clip and consumables in the pre-cooled converter;
2. Activate PVDF membrane with methanol for 1 min and rinse with transfer buffer;
3. Install it in the order of "black edge of clip - sponge - filter paper - filter paper - glue -PVDF membrane - filter paper - filter paper - sponge - white edge of clip";
4. The protein was electrotransferred to PVDF membrane. (0.45 µm PVDF membrane is recommended) Reference Table for Selecting PVDF Membrane Pore Size Specifications
Recommended conditions for wet transfer: **200 mA, 120 min**.
(Note that the transfer conditions can be adjusted according to the protein size. For high-molecular-weight proteins, a higher current and longer transfer time are recommended. However, ensure that the transfer tank remains at a low temperature to prevent gel melting.)

Block
1. After electrotransfer, wash the film with TBST at room temperature for 5 minutes;
2. Incubate the film in the blocking solution for 1 hour at room temperature;
3. Wash the film with TBST for 3 times, 5 minutes each time.

Antibody incubation
1. Use 5% skim milk powder to prepare the primary antibody working liquid (recommended dilution ratio for primary antibody 1:1000), gently shake and incubate with the film at 4°C overnight;
2. Wash the film with TBST 3 times, 5 minutes each time;
3. Add the secondary antibody to the blocking solution and incubate with the film gently at room temperature for 1 hour;
4. After incubation, wash the film with TBST 3 times for 5 minutes each time.

Antibody staining
1389. Add the prepared ECL luminescent substrate (or select other color developing substrate according to the second antibody) and mix evenly;
2. Incubate with the film for 1 minute, remove excess substrate (keep the film moist), wrap with plastic film, and expose in the imaging system. **(Exposure time of at least 60s is recommended)**

References

https://pubmed.ncbi.nlm.nih.gov/8552098/ https://pubmed.ncbi.nlm.nih.gov/24438093/

Application Data

WB

Validated by Capsid Bio



Lane 1: SK-N-MC
Lane 2: SK-N-MC (Forskolin- and FGF-treated)

IF

Validated by Capsid Bio



Immunofluorescent analysis of Hela cells using F0154 (green, 1:400), Hoechst (blue) and tubulin (Red).