# Product Datasheet

## Biological Description

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| Specificity | Phospho-c-Myc (Thr58) Antibody [L1K9] detects endogenous levels of total c-Myc protein only when it is phosphorylated at Thr58. |
| Background | Phospho-c-Myc Thr58 refers to the threonine-58 phosphorylated form of c-Myc, which is a proto-oncogenic basic helix-loop-helix leucine zipper transcription factor at the core of the Myc/Max/Mad network, regulating a significant portion of the genome involved in cell proliferation, metabolism, and apoptosis. c-Myc comprises an N-terminal transactivation domain containing Myc box I and II with Thr58 and Ser62 residues, a central HLH-LZ motif for heterodimerization with Max and binding to E-box DNA sequences, and a C-terminal region with degradation signals. Phosphorylation at Thr58, which is primed by prior phosphorylation of Ser62 by ERK or CDKs, is mediated by GSK-3 beta and creates a site for dephosphorylation at Ser62 by PP2A. This event recruits the SCF-Fbw7 E3 ligase, leading to K48-linked ubiquitination at lysines such as Lys57 and Lys161 and subsequent proteasomal degradation of c-Myc, resulting in a short protein half-life of about 20 to 30 minutes. The presence of phospho-Thr58 destabilizes c-Myc, limits its transcriptional activation of growth-related genes like nucleolin, CDK4, and hTERT, and influences its subnuclear localization to promyelocytic leukemia bodies with GSK-3, ultimately reducing its oncogenic potential. This phosphorylation enforces rapid turnover of Myc, maintaining homeostatic control during mitogen signaling. phospho-Thr58 helps balance Ras and ERK-driven proliferation with protein degradation checkpoints during the G1 to S phase transition and spindle assembly. Mutations at Thr58, such as T58A, prevent phosphorylation, stabilize c-Myc, enhance cellular transformation, and result in genomic instability due to dysregulation of the spindle assembly checkpoint and upregulation of Aurora B. Loss or absence of phospho-Thr58 is frequently observed in Burkitt lymphoma, where it represents a common mutational hotspot, as well as in other MYC-driven cancers including breast and prostate cancer, and is associated with poor prognosis due to uncontrolled Myc activity. |

## Usage Information

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| Application | WB, IF, FCM | Dilution | WB: 1:1000; IF: 1:250; FCM: 1:700 |
| Reactivity | Human | | |
| Source | Rabbit Monoclonal Antibody | MW | 48 kDa |
| Storage Buffer | PBS, pH 7.2+50% Glycerol+0.05% BSA+0.01% NaN3 | | |
| Storage (from the date of receipt) | -20°C (avoid freeze-thaw cycles), 2 years | | |

## Experimental Methods

**WB**

Experimental Protocol:

Sample preparation
1. Tissue: Lyse the tissue sample by adding an appropriate volume of ice-cold **RIPA/NP-40 Lysis Buffer (containing Protease Inhibitor Cocktail, Phosphatase Inhibitor Cocktail)**，and homogenize the tissue at a low temperature or lyse it by sonication on ice, then incubate on ice for 30 minutes.
2. Adherent cell: Aspirate the culture medium and wash the cells with ice-cold PBS twice. Lyse the cells by adding an appropriate volume of **RIPA/NP-40 Lysis Buffer (containing Protease Inhibitor Cocktail, Phosphatase Inhibitor Cocktail)**, sonicate to lyse the cells, and incubate on ice for 30 minutes.
3. Suspension cell: Transfer the culture medium to a pre-cooled centrifuge tube. Centrifuge and aspirate the supernatant. Wash the cells with ice-cold PBS twice. Lyse the cells by adding an appropriate volume of **RIPA/NP-40 Lysis Buffer (containing Protease Inhibitor Cocktail, Phosphatase Inhibitor Cocktail)**, sonicate to lyse the cells, and incubate on ice for 30 minutes.
4. Place the lysate into a pre-cooled microcentrifuge tube. Centrifuge at 4°C for 15 min. Collect the supernatant;
5. Remove a small volume of lysate to determine the protein concentration;
6. Combine the lysate with protein loading buffer. Boil 20 µL sample under 95-100°C for 5 min. Centrifuge for 5 min after cool down on ice.

Electrophoretic separation
1. According to the concentration of extracted protein, load appropriate amount of protein sample and marker onto SDS-PAGE gels for electrophoresis. Recommended separating gel (lower gel) concentration: 10%. Reference Table for Selecting SDS-PAGE Separation Gel Concentrations
2. Power up 80V for 30 minutes. Then the power supply is adjusted (110 V~150 V), the Marker is observed, and the electrophoresis can be stopped when the indicator band of the predyed protein Marker where the protein is located is properly separated. (Note that the current should not be too large when electrophoresis, too large current (more than 150 mA) will cause the temperature to rise, affecting the result of running glue. If high currents cannot be avoided, an ice bath can be used to cool the bath.)

Transfer membrane
1. Take out the converter, soak the clip and consumables in the pre-cooled converter;
2. Activate PVDF membrane with methanol for 1 min and rinse with transfer buffer;
3. Install it in the order of "black edge of clip - sponge - filter paper - filter paper - glue -PVDF membrane - filter paper - filter paper - sponge - white edge of clip";
4. The protein was electrotransferred to PVDF membrane. ( 0.45 µm PVDF membrane is recommended ) Reference Table for Selecting PVDF Membrane Pore Size Specifications
Recommended conditions for wet transfer: **200 mA, 120 min**.
( Note that the transfer conditions can be adjusted according to the protein size. For high-molecular-weight proteins, a higher current and longer transfer time are recommended. However, ensure that the transfer tank remains at a low temperature to prevent gel melting.)

Block
1. After electrotransfer, wash the film with TBST at room temperature for 5 minutes;
2. Incubate the film in the blocking solution **( recommending 5% BSA solution)** for 1 hour at room temperature;
3. Wash the film with TBST for 3 times, 5 minutes each time.

Antibody incubation
1. Use 5% skim milk powder to prepare the primary antibody working liquid (recommended dilution ratio for primary antibody 1:1000), gently shake and incubate with the film at 4°C overnight;
2. Wash the film with TBST 3 times, 5 minutes each time;
3. Add the secondary antibody to the blocking solution and incubate with the film gently at room temperature for 1 hour;
4. After incubation, wash the film with TBST 3 times for 5 minutes each time.

Antibody staining
1. Add the prepared ECL luminescent substrate (or select other color developing substrate according to the second antibody) and mix evenly;
2. Incubate with the film for 1 minute, remove excess substrate (keep the film moist), wrap with plastic film, and expose in the imaging system.

**IF**

Experimental Protocol:

**Sample Preparation**
1. Adherent Cells: Place a clean, sterile coverslip in a culture dish. Once the cells grow to near confluence as a monolayer, remove the coverslip for further use.
2. Suspension Cells: Seed the cells onto a clean, sterile slide coated with poly-L-lysine.

## Experimental Methods

3. Frozen Sections: Allow the slide to thaw at room temperature. Wash it with pure water or PBS for 2 times, 3 minutes each time.
4. Paraffin Sections: Deparaffinization and rehydration. Wash the slide with pure water or PBS for 3 times, 3 minutes each time. Then perform antigen retrieval.

**Fixation**
1. Fix the cell coverslips/spots or tissue sections at room temperature using a fixative such as 4% paraformaldehyde (4% PFA) for 10-15 minutes.
2. Wash the sample with PBS for 3 times, 3 minutes each time.

**Permeabilization**
1.Add a detergent such as 0.1–0.3% Triton X-100 to the sample and incubate at room temperature for 10–20 minutes.
(Note: This step is only required for intracellular antigens. For antigens expressed on the cell membrane, this step is unnecessary.)
Wash the sample with PBS for 3 times, 3 minutes each time.

**Blocking**
Add blocking solution and incubate at room temperature for at least 1 hour. (Common blocking solutions include: serum from the same source as the secondary antibody, BSA, or goat serum.)
**Note: Ensure the sample remains moist during and after the blocking step to prevent drying, which can lead to high background.**

**Immunofluorescence Staining (Day 1)**
1. Remove the blocking solution and add the diluted primary antibody.
2. Incubate the sample in a humidified chamber at 4°C overnight.

**Immunofluorescence Staining (Day 2)**
1. Remove the primary antibody and wash with PBST for 3 times, 5 minutes each time.
2. Add the diluted fluorescent secondary antibody and incubate in the dark at 4°C for 1–2 hours.
3. Remove the secondary antibody and wash with PBST for 3 times, 5 minutes each time.
4. Add diluted DAPI and incubate at room temperature in the dark for 5–10 minutes.
5. Wash with PBST for 3 times, 5 minutes each time.

**Mounting**
1. Mount the sample with an anti-fade mounting medium.
2. Allow the slide to dry at room temperature overnight in the dark.
3. Store the slide in a slide storage box at 4°C, protected from light.

## References

https://pubmed.ncbi.nlm.nih.gov/14563837/ https://pubmed.ncbi.nlm.nih.gov/15103331/

## Application Data

### IF

Validated by Capsid Bio




Immunofluorescent analysis of Hela cells using F2474 (green, 1:250), Hoechst (blue) and tubulin (Red).

### WB

Validated by Capsid Bio




Lane 1: HeLa, Lane 2: HeLa (Lambda Phosphatase-treated)