Phospho-HDAC4 (S246)/5 (S259)/7 (S155) Antibody [H5N6]

Catalog No.C18793 Batch:C1879303

Product Datasheet

Biological Description

Specificity	Phospho-HDAC4 (S246)/5 (S259)/7 (S155) Antibody [H5N6] detects endogenous levels of HDAC4, HDAC5 and HDAC7 proteins only when phosphorylated on Ser246, Ser259 and Ser155, respectively.
Background	Phospho-HDAC4 (S246), HDAC5 (S259), and HDAC7 (S155) denote the activated, cytoplasmic forms of these class IIa histone deacetylases, which feature an N-terminal regulatory domain with conserved serine phosphorylation sites, S246 in HDAC4, S259 in HDAC5, and S155 in HDAC7, for 14-3-3 chaperone binding, and a C-terminal catalytic domain containing a zinc-dependent active site within a conserved HDAC fold of parallel β-sheets and flanking α-helices for substrate recognition and deacetylation. These structural motifs facilitate nuclear-cytoplasmic shuttling: kinases such as CaMK and PKD phosphorylate the conserved serines in response to signals like calcium influx, VEGF, or T-cell activation, prompting 14-3-3 binding and nuclear export, which relieves repression of MEF2-driven genes critical for muscle differentiation, endothelial angiogenesis, lymphocyte development, and neuronal survival. HDAC4/5/7 recruit N-CoR/BCoR corepressors to deacetylate histone and non-histone substrates, compacting chromatin and suppressing transcription until signal-induced phosphorylation leads to their cytoplasmic sequestration and derepression of key pathways; dysregulation that sustains nuclear retention of these HDACs is implicated in diseases such as lymphomas or cardiovascular disorders, driving pathological proliferation and fibrosis through unchecked ERK or NFAT signaling.

Usage Information

Application	WB	Dilution	
Reactivity	Human, Mouse		
Source	Rabbit Monoclonal Antibody	MW	140 kDa, 124 kDa
Storage Buffer	PBS, pH 7.2+50% Glycerol+0.05% BSA+0.01% NaN3		
Storage (from the date of receipt)	-20°C (avoid freeze-thaw cycles), 2 years		

Experimental Methods

WB

Experimental Protocol:

Sample preparation
1. Tissue: Lyse the tissue sample by adding an appropriate volume of ice-cold **RIPA/NP-40 Lysis Buffer (containing Protease Inhibitor Cocktail, Phosphatase Inhibitor Cocktail)**，and homogenize the tissue at a low temperature or lyse it by sonication on ice, then incubate on ice for 30 minutes.
2. Adherent cell: Aspirate the culture medium and wash the cells with ice-cold PBS twice. Lyse the cells by adding an appropriate volume of **RIPA/NP-40 Lysis Buffer (containing Protease Inhibitor Cocktail, Phosphatase Inhibitor Cocktail)**, sonicate to lyse the cells, and incubate on ice for 30 minutes.
3. Suspension cell: Transfer the culture medium to a pre-cooled centrifuge tube. Centrifuge and aspirate the supernatant. Wash the cells with ice-cold PBS twice. Lyse the cells by adding an appropriate volume of **RIPA/NP-40 Lysis Buffer (containing Protease Inhibitor Cocktail, Phosphatase Inhibitor Cocktail)**, sonicate to lyse the cells, and incubate on ice for 30 minutes.
4. Place the lysate into a pre-cooled microcentrifuge tube. Centrifuge at 4°C for 15 min. Collect the supernatant;
5. Remove a small volume of lysate to determine the protein concentration;
6. Combine the lysate with protein loading buffer. Boil 20 µL sample under 95-100°C for 5 min. Centrifuge for 5 min after cool down on ice.

Electrophoretic separation
1. According to the concentration of extracted protein, load appropriate amount of protein sample and marker onto SDS-PAGE gels for electrophoresis. **Recommended separating gel (lower gel) concentration: 5%.** Reference Table for Selecting SDS-PAGE Separation Gel Concentrations
2. Power up 80V for 30 minutes. Then the power supply is adjusted (110 V~150 V), the Marker is observed, and the electrophoresis can be stopped when the indicator band of the predyed protein Marker where the protein is located is properly separated. (Note that the current should not be too large when electrophoresis, too large current (more than 150 mA) will cause the temperature to rise, affecting the result of running glue. If high currents cannot be avoided, an ice bath can be used to cool the bath.)

Transfer membrane
1. Take out the converter, soak the clip and consumables in the pre-cooled converter;
2. Activate PVDF membrane with methanol for 1 min and rinse with transfer buffer;
3. Install it in the order of "black edge of clip - sponge - filter paper - filter paper - glue -PVDF membrane - filter paper - filter paper - sponge - white edge of clip";
4. The protein was electrotransferred to PVDF membrane. (0.45 µm PVDF membrane is recommended) Reference Table for Selecting PVDF Membrane Pore Size Specifications
Recommended conditions for wet transfer: **200 mA, 120 min**.
(Note that the transfer conditions can be adjusted according to the protein size. For high-molecular-weight proteins, a higher current and longer transfer time are recommended. However, ensure that the transfer tank remains at a low temperature to prevent gel melting.)

Block
1. After electrotransfer, wash the film with TBST at room temperature for 5 minutes;
2. Incubate the film in the blocking solution **(recommending 5% BSA solution)** for 1 hour at room temperature;
3. Wash the film with TBST for 3 times, 5 minutes each time.

Antibody incubation
1. Use 5% skim milk powder to prepare the primary antibody working liquid (recommended dilution ratio for primary antibody 1:1000), gently shake and incubate with the film at 4°C overnight;
2. Wash the film with TBST 3 times, 5 minutes each time;
3. Add the secondary antibody to the blocking solution and incubate with the film gently at room temperature for 1 hour;
4. After incubation, wash the film with TBST 3 times for 5 minutes each time.

Antibody staining
1. Add the prepared ECL luminescent substrate (or select other color developing substrate according to the second antibody) and mix evenly;
2. Incubate with the film for 1 minute, remove excess substrate (keep the film moist), wrap with plastic film, and expose in the imaging system.

References

https://pubmed.ncbi.nlm.nih.gov/10869435/ https://pubmed.ncbi.nlm.nih.gov/25616866/

Application Data

WB

Validated by Capsid Bio



Lane 1: HeLa, Lane 2: HeLa (phosphatase treated), Lane 3: NIH/3T3, Lane 4: NIH/3T3 (phosphatase treated)